

The worldwide blood parasite: malaria

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Human malaria is a worldwide problem. More than one-third of the world's population (about two billion people) live in malaria-endemic areas, and one billion people are estimated to be carriers of the causative parasite, *Plasmodium* spp., at any time.¹ About 500 million people become infected annually, resulting in more than two million deaths.² *Plasmodium falciparum* is the main cause of severe malaria outbreaks and mortality rate. Moreover, the increasing problem of multidrug resistant strains necessitates the discovery of new and highly efficacious drugs.

S-Adenosyl-L-homocysteine (SAH) hydrolase has emerged as a target enzyme for the molecular design of antiviral, antitumor, antiparasitic, antiarthritic and immunosuppressive agents.^{3,4,5} SAH is formed after the donation of the methyl group of S-adenosyl-L-methionine (SAM) to a methyl acceptor and is hydrolyzed to adenosine and homocysteine by SAH hydrolase, physiologically. Inhibition of SAH hydrolase results in cellular accumulation of SAH. It is a potent feedback inhibitor of SAM-dependent biological methylation such as the 5'-end of eukaryotic mRNA.^{3,6} In contrast to human SAH hydrolase, *P. falciparum* SAH hydrolase contains a 41-amino acid insert (Gly145-Lys185) inside the sequence.⁷ *P. falciparum* causes malignant malaria. This difference may produce selective sensitivity against each SAH hydrolase inhibitor. Neplanocin A and aristeromycin are naturally occurring products possessing inhibitory activity against SAH hydrolase. When these inhibitors work as a substrate for adenosine kinase, they show cytotoxicity. Neplanocin A and aristeromycin are also known to be rapidly deaminated by adenosine deaminase to a chemotherapeutically inactive inosine congener.⁸ In order to overcome these disadvantages in the development of chemotherapeutic agents, chemical modifications of carbocyclic nucleosides have been carried out. Because noraristeromycin lacks the 5'-methylene unit of aristeromycin, it does not work as a substrate for adenosine deaminase.

We have found that the introduction of fluorine to the 2-position of noraristeromycin increased selective inhibition against *P. falciparum* SAH (PfSAHH) compared with human SAH (HsSAHH).⁹ PfSAHH has additional space near the 2-position of the adenine-ring, in the substrate binding pocket compared with HsSAHH.¹⁰ Mutagenic analysis of the amino acid residue forming the additional space confirmed that inhibitor selectivity is due to the difference of only one amino acid residue.¹¹ We carried out synthesis of several kinds of carbocyclic nucleosides and evaluated their inhibitory activity against PfSAH and HsSAHH.¹² Recently, we have found *P. falciparum* TMP kinase as a new target enzyme for the development of anti-malaria agents.¹³ Therefore, the properties of malaria TMP kinase and synthesis of its inhibitors are also

discussed. These results will significantly contribute to the design of anti-malaria agents.

Human malaria is a worldwide problem and more than one-third of the world population (about two billion peoples) live in malaria-endemic areas, and a billion people are estimated to be carriers of the causative parasite, *Plasmodium* spp., at any one time.¹ About 500 million people become infected annually, resulting in more than 2 million deaths.² *Plasmodium falciparum* is the main cause of severe malaria outbreaks and high deaths. Moreover, the increasing problem of multidrug resistant strains necessitates the discovery of new, highly efficacious drugs. Figure 1 shows epidemic area of malaria and a conventional anti-malaria drugs, such as chloroquine, quinine, proguanil, and artemisinin. Malaria currently spreading tolerate a conventional anti-malaria drug such as chloroquine. Therefore, we are currently developing new drugs which will be more effective in controlling the spread of malaria, (Figure 1). S-Adenosyl-L-homocysteine (SAH) hydrolase has emerged as a target enzyme for the molecular design of antiviral, antitumor, antiparasitic, antiarthritic, and immunosuppressive agents.^{3,4,5} SAH is formed after the donation of the methyl group of S-adenosyl-L-methionine (SAM) to a methyl acceptor and is hydrolyzed to adenosine and homocysteine by SAH hydrolase, physiologically. Inhibition of SAH hydrolase results in cellular accumulation of SAH. It is a potent feedback inhibitor of SAM-dependent biological methylation such as the 5'-end eukaryotic mRNA.^{3,6} (Figure 2).

Palmer and Abeles have elucidated the mechanism of SAH hydrolase. In the hydrolytic direction, the first step involves oxidation of the 3'-hydroxyl group of SAH by enzyme-bound NAD⁺ followed by β-elimination of homocysteine to give the 3'-keto-4',5'-didehydro-5'-deoxyadenosine as an enzymatically bound intermediate. Michael's addition of water to this intermediate affords the 3'-ketoadenosine, which is then reduced by enzyme-bound NADH to produce adenosine.

SAH hydrolase can also catalyze the reaction in the synthetic direction via the same mechanism, using adenosine and homocysteine as substrates and affording SAH, (Figure 3).

In contrast to human SAH hydrolase, *P. falciparum* SAH hydrolase contains a 15 amino acid insert (Gly145 - Lys185) inside the sequence.⁷ *P. falciparum* causes malignant malaria. This difference may produce selective sensitivity against each SAH hydrolase inhibitor, (Figure 4).

Figure 5 shows SAH hydrolase inhibitors. These analogues included adenosine analogues with acyclic "sugar" moieties such as (S)-DHPA and D-eritadenine. Neplanocin A and aristeromycin are natural products. These compounds are quite potent. Unfortunately, the cytotoxicity of these compounds precludes their clinical use as antiviral agents. Clearly, removal of the hydroxymethyl substituent would preclude 5'-phosphorylation by adenosine kinase. Therefore, DHCa and noraristeromycin were designed. These compounds retain antiviral activity while their toxicity is considerably lower than the parent compound. When these inhibitors work as a substrate for adenosine kinase, they show low cytotoxicity. Neplanocin A and aristeromycin are also known to be rapidly deaminated by adenosine deaminase to a chemotherapeutically inactive inosine congener.⁸ On the other hand, a 4',5'-unsaturated 5'-fluoroadenosine (M

28,842), an inhibitor of SAH hydrolase, was found to inhibit markedly the growth of *Plasmodium falciparum* in vitro. SAH hydrolase represents a worthwhile target for the development of anti-malaria agents, (Figure 5).

In order to overcome these disadvantages in the development of chemotherapeutic agents, chemical modifications of carbocyclic nucleosides have been carried out. Because noraristeromycin lacks the 5'-methylene unit of aristeromycin, it does not work as a substrate for adenosine deaminase.

Figure 4. Selective inhibition against *P. falciparum* SAH hydrolase

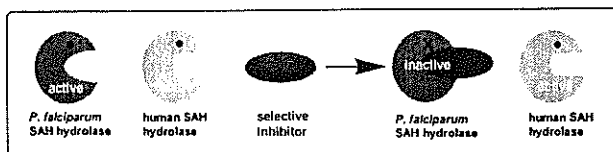
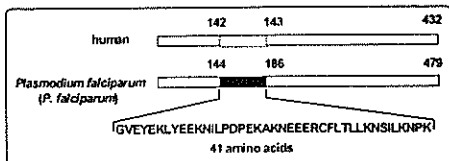


Figure 5. SAH hydrolase Inhibitors

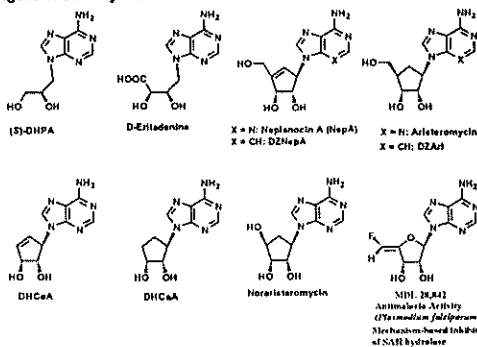


Table 1

human SAH hydrolase IC ₅₀ , μM	560	76	1.1	60
<i>P. falciparum</i> SAH hydrolase IC ₅₀ , μM	60	7.9	3.1	18

Y. Kitade, A. Kozaki and C. Yotome. *Tetrahedron Lett.*, 42, 433-435 (2001)

Y. Kitade, A. Kozaki, Y. Miwa and M. Nakanishi. *Tetrahedron*, 58, 1271-1277 (2002)

Figure 7. Synthesis of 2-fluoronoraristeromycin

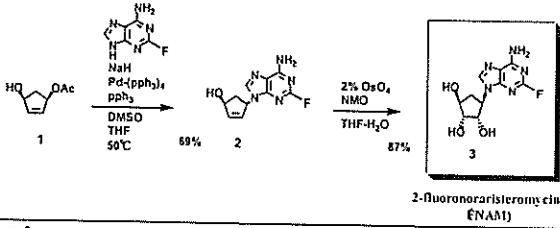
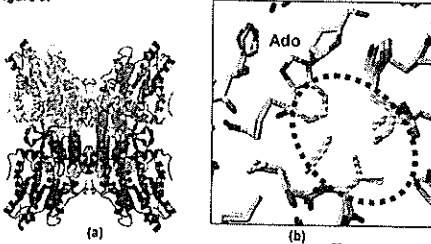


Figure 8.



T60 (human)
C59 (*P. fa*)

Figure 9.

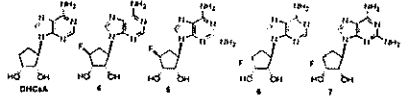
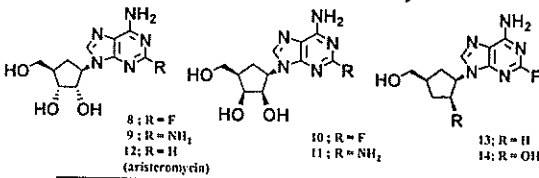


Figure 10. Inhibitory activities 2-modified aristeromycins



compound	IC ₅₀ (µM)		Selective Index
	Hs SAHH	Pf SAHH	
8	47.2	1.98	24
9	90.7	4.51	20
12	4.85	57	0.085
10,11,13,14	>500	>500	-

280-fold increased

Selective index : IC₅₀ of Hs SAHH/ IC₅₀ of Pf SAHH

Ando, T., Iwata, M., Zulfqar, F., Miyamoto, T., Nakanishi, M., & Kitade, Y., *Bioorg. & Med. Chem.*, 16, 3809-3815 (2008).

Figure 11. Resistance against adenosine deaminase

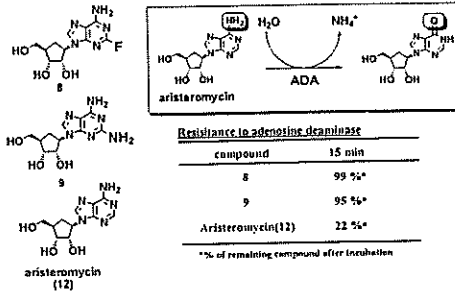


Figure 12. Pyrimidine metabolism

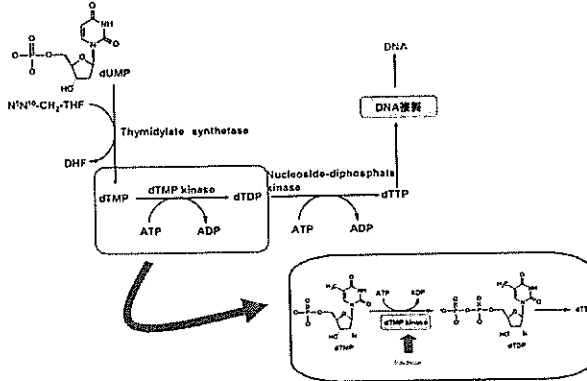
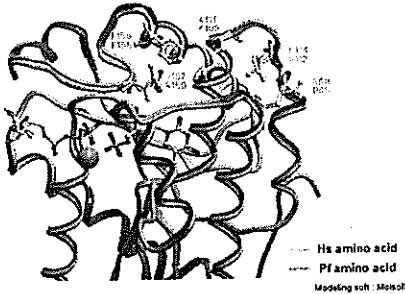


Figure 13. Modeling of Hs TMP kinase and Pf TMP kinase



We prepared modified carbocyclic purine analogs as shown in Figure 6. Along the top row as shown in Table 1, I have listed IC₅₀ values against human SAH hydrolase and the bottom row shows IC₅₀ values against P.falciparum SAH hydrolase. The smaller number, the greater the activity. Among these derivatives, 2-amino-noraristeromycin selectively showed inhibitory activity against P. falciparum SAH hydrolase. Therefore, we envisaged that introduction

of a fluorine atom to the 2-position of the adenine ring could increase selectivity against *P. falciparum* SAH hydrolase.

At first, we prepared 2-fluoronoraristeromycin (3) as shown in Figure 7. The palladium-coupling reaction of compound (1) with sodium salt of commercially available 2-fluoroadenine resulted in the formation of the coupling compound (2) in 69 % yield. Subsequent oxidation of compound 2 was carried out with osmium tetroxide (OsO₄) in the presence of 4-methylmorpholine N-oxide (NMO) to give 2-fluoronoraristeromycin (3) in 87 % yield. These structures of compounds 2 and 3 were supported by spectral data and microanalytical results.

A profile of the inhibitory activity of 2-fluoronoraristeromycin (3; FNAM) against *P. falciparum* and human SAH hydrolase is shown in Table 2. Introduction of a fluorine atom to the adenine ring causes selectivity against *P. falciparum* SAH hydrolase. The K_i values of FNAM against human and *P. falciparum* SAH hydrolase were 7.9 μM and 0.48 μM, respectively. The selective index was 16. The introduction of a fluorine atom brought an 18-fold increase in the selective index. In vitro antimalaria activity and cytotoxicity are shown in Table 3. The cytotoxicity of the 2-fluoro derivative (3) against FM3A cells brought about a 100% decrease. However, antimalaria activity of 2-fluoronoraristeromycin (3) was retained.

Isolation of non-fusion malaria enzyme, *P. falciparum* SAH hydrolase, has succeeded with difficulty. But isolation of His-tag malaria enzyme gave a large amount of protein with high purity. This protein is applicable to X-ray crystallography analysis. Finally, we were succeeding the crystallization of malaria SAH hydrolase. We spent three years for the crystallization of *P. falciparum* SAH hydrolase. We analyzed the structure of malaria SAH hydrolase using Spring-8 at Harima, Japan. The crystallographic analysis revealed the structure of malaria SAH hydrolase as shown Figure 8 (a). The enzyme, in its active form, is a homotrimer of identical subunits and has a molecular mass of approximately 100 kDa. The insertion sequence of 41-amino acids is far from the active site. Figure 8 (b) shows the adenosine binding site of malaria SAH hydrolase. Interestingly, we have found a crucial structural difference between the adenosine binding site of malaria enzyme and that of human enzyme. Thr60 located near the C2 of the adenine ring of the adenosine in human enzyme. In malaria enzyme, Thr60 is replaced by Cys59. The malaria SAH hydrolase has more space for a fluorine atom at the 2-position of 2-fluoroadenine carbocyclic nucleosides. This observation explains the selectivity of 2-fluoroadenine derivatives against malaria SAH hydrolase. We have found that the introduction of a fluorine to the 2-position of noraristeromycin increases selective inhibition against *P. falciparum* SAH hydrolase (PfSAHH) compared with human SAH hydrolase (HsSAHH).⁹ PfSAHH has additional space near the 2-position of the adenine-ring, in the substrate binding pocket compared with HsSAHH.¹⁰ Mutagenic analysis of the amino acid residue forming the additional space confirmed that inhibitor selectivity is due to the difference of only one amino acid residue.¹¹ (Figure 8).

We carried out synthesis of several kinds of carbocyclic nucleosides and evaluated their inhibitory activity against PfSAHH and HsSAHH.¹² 4'-Fluoro derivative 4 showed inhibitory activity against human enzyme. On the other hand, the 2-amino 4'-fluoro derivative (7) showed inhibitory activity against

malaria enzyme. These results will contribute greatly to the design of potent inhibitors against *P. falciparum* SAH hydrolase, (Figure 9 & Table 4)

The inhibitory activities of base- and/or sugar -modified aristeromycins against HsSAHH and PfSAHH are summarized as IC₅₀ values in Table of Figure 10. The 2-fluorinated aristeromycin (8) and 2-amino compound 9 showed strong inhibitory activities against PfSAHH with IC₅₀ value of 1.98 and 4.51 μ M and superior selective index 24 and 20, respectively. These values were better than those of the typical compounds which we had synthesized. The other synthetic compounds did not show any inhibitory activity up to 500 μ M. The susceptibility of these compounds to adenosine deaminase was investigated under the previously reported conditions as show in Figure 11. 2-Fluoroaristeromycin 8 was completely resistant to adenosine deaminase unlike aristeromycin (12), which was rapidly deaminated within 15 min under the same reaction conditions. 2-Aminoaristeromycin 9 was slightly deaminated with a 5% conversion ratio. The introduction of a halogen atom was effective in designing inhibitors showing the resistance to adenosine deaminase. This observation will significantly contribute to the design of potent inhibitors as anti-malarial agents. (See Figure10 & 11).

Recently, we have found *P. falciparum* TMP kinase as a new target enzyme for the development of anti-malaria agents.¹³ Plasmodium falciparum thymidylate kinase (PFTMK) can tolerate a range of substrates, which distinguishes it from other thymidylate kinases. The enzyme not only phosphorylates TMP and dUMP but can also tolerate bulkier purines, namely, dGMP, GMP, and dIMP. In order to probe the flexibility of PFTMK in accommodating ligands of various sizes, we developed six mutant enzymes and subjected to thermodynamic, inhibitory and catalytic evaluation. Mutation of Phe74 to alanine equally affected TMP and dGMP kinase activity. Kinase activity was markedly affected by introducing a larger lysine residue instead of A111. The lack of the hydroxyl group after inducing mutation of Y107F affected enzyme activity, and had more severe impact on dGMP kinase activity. PFTMK can be inhibited by both purine and pyrimidine nucleosides, raising the possibility of developing highly selective drugs. Thermodynamic analysis revealed that enthalpic forces govern both purine and pyrimidine nucleoside monophosphate binding, and the binding affinity of both substrates was highly comparable. The heat produced due to dGMP binding is lower than that attributable to TMP. This indicates that additional interactions occur with TMP, which may be lost with larger dGMP. Targeting PFTMK not only affects thymidine nucleotide synthesis but may also affect purine nucleotides, and thus it has become an attractive antimicrobial target. This study provides differentiations between some similarities and differences in the interactions of PFTMK with TMP and dGMP. In this context, similar properties and variations provide valuable information for optimizing new selective inhibitors. Therefore, the properties of malaria TMP kinase and synthesis of its inhibitors are also discussed, (Figure 12 and 13)

These results described in this paper will significantly contribute to the design of anti-malarial agents.

References

Guerin, P.J., Olliaro, P., Nosten, F., Druilhe, P., Laxminarayan, R., Binka, F., Kilama, W.L., Ford, F.

- and White, N.J., *The Lancet Infectious Diseases*, 2002, 2, 564-573.
- Breman, J.G., Egan, A. and Keusch, G.T., *Am. J. Trop. Med. Hyg.*, 2001, 64, 4-7.
- (a) Keller, B. T.; Borchardt, R. T. In *Biological Methylation and Drug design: Experimental and Clinical Roles of S-adenosylmethionine*, Borchardt, R. T., Creveling, C. R., Ueland, P. M., E Humana: Clifton, NJ, 1986; pp.385. (b) Wolfe, M. S.; Borchardt, R. T., *J. Med. Chem.* 1984, 27, 1521- 1530. (c) Chiang, P. K., *Pharmacol. Ther.* 1998, 77, 115-134 and references therein.
- Wolos, J. A.; Frondorf, K. A.; Babcock, G. F.; Stripp, S. A.; Bowlin, T. L., *Cell. Immunol.*, 1993, 147, 402-408.
- (a) Bitonti, A. J.; Baumann, R. J.; McCann, P. P., *Biochem. Pharmacol.*, 1990, 40, 601–606. Henderson, D.M.; Hanson, S.; Allen, T.; Wilson, K.; Coulter-Karis, D. E.; Grennberg, M. Hershfield, M. S.; Ullman, B., *Mol. Biochem. Parasitol.*, 1992, 53, 169–183. (c) Wolos A.; Frondorf, K. A.; Esser, R. E., *J. Immunol.*, 1993, 151, 526–534.
- Liu, S.; Wolfe, M. S.; Borchardt, R. T., *Antiviral Res.*, 1992, 19, 247-265.
- Creedon, K. A.; Rathod, P. K.; Wellems, T. E., *J. Biol. Chem.*, 1994, 269, 16364-16370.
- Keller, B. T.; Borchardt, R. T. In *Biological Methylation and Drug Design*; Borchardt, R. T.; Creveling, C. R.; Ueland, P. M.; Eds.; Humana: Clifton, NJ; 1987, 385-396.
- Kitade, Y.; Kojima, H.; Zulfiquar, F.; Kim, H. -S.; Wataya, Y. *Bioorg. Med. Chem. Lett.* 2003, 13, 39
- Tanaka, N.; Nakanishi, M.; Kusakabe, Y.; Shiraiwa, K.; Yabe, S.; Ito, Y.; Kitade, Y.; Nakamura, K. *Mol. Biol.* 2004, 343, 1007.
- Nakanishi, M.; Yabe, S.; Tanaka, N.; Ito, Y.; Nakamura, K. T.; Kitade, Y. *Mol. Biochem. Parasitol.* 2005, 143, 146.
- Nakanishi, M.; Iwata, A.; Yatome, C.; Kitade, Y., *J. Biochem.*, 2001, 129, 101-105; Kitade, Kozaki, A.; Miwa, T.; Nakanishi, M., *Tetrahedron.*, 2002, 58, 1271-1277; Kitade, Y.; Ando Yamaguchi, T.; Hori, A.; Nakanishi, M.; Ueno, Y. *Bioorg. Med. Chem.* 2006, 14, 5578-5585
- Ando, T.; Iwata, M.; Zulfiquar, F.; Miyamoto, T.; Nakanishi, M.; Kitade, Y., *Bioorg. Med. Chem.* 2008, 16, 3809-3815; Kojima, H., Kozaki, A., Iwata, M., Ando, T., and Kitade, Y., *Bio Med. Chem.*, 2008, 16, 6575-6579; Zulfiquar, F., Kojima, H., Nakanishi, M., Ando, T., Kitade, Y., *Nucleosides Nucleotides Nucleic Acids*, 2008, 27, 1153-1157.
- Kandeel, M. and Kitade, Y., *J. Biochem.*, 144, 245-250 (2008); Kandeel, M., Ando, T., Kitamura, Abdel-Aziz, M., and Kitade, Y., *Parasitology*, 136, 11-25 (2009); Kandeel, M., Miyamoto and Kitade, Y., *Biol. Pharm. Bull.*, 32, 1321-1327 (2009).