

Clinical and Etiological study on respiratory affections of sheep

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Abstract

The results of this study illustrate that the respiratory affections of sheep occur in two forms the first one was sever with acute signs while the other mild with prolonged cough and loss of body condition. The bacteria associated with such affections were isolated in single and mixed form. The most prevalent isolated bacteria was Mannheimia haemolytica that also associated with sever and acute signs. E.coli, Pasteurella multocida, Staphylococcus aureus, Klebsiella pneumoniae, Pseudomonas aeruginosa, Streptococcus pneumoniae and Salmonella were isolated at variable percentages. The antibiogram and treatment trials revealed that the most effective antimicrobial agents against these bacteria were flurophenicol and enrofloxacin but oxytetracycline shown reduced activity in both antibiogram and treatment trials.

Key words Sheep, Respiratory affection, Mannheimia haemolytica, Treatment.

Introduction:

Respiratory affections occur frequently in sheep. In many countries respiratory diseases represent the most serious sheep problem and can be an important cause of death and reduced productivity (Martin, 1996). Respiratory disorder appears to be a complex disease. Many infectious agents synchronized with stress and/or environmental factors responsible for such disease causing a considerable level of economic losses of the infected flocks (Alley, 1975; Al-darraj et al., 1982 and Sharma and Woldehiwet, 1991). The infectious agents associated with respiratory affections of sheep include viral agents such as PI3 virus, Reo virus, Respiratory syncytial virus and Ovine adenovirus type 6 (Sharp and Nettleton, 2000). Mycoplasma ovipneumoniae and Mycoplasma argini also incriminated as cause of respiratory disease condition in sheep. The viral and mycoplasmal agents causing low grade respiratory disease with mild signs after extended period of infection (Martin, 1996). Those which cause mortalities and obvious acute clinical signs are associated with bacterial agents particularly Mannheimia haemolytica (Donachie, 2000).

The fundamental goal of the present work carried out to:

- 1-Clinical description of the respiratory disorder manifestation in sheep.
- 2-Isolation and identification of the possible causative bacterial pathogens from diseased animals.
- 3- Antibiogram of the isolated bacterial agents, and treatment trials of some forms of the disease with different drugs to reaching the most suitable one.

Materials and Methods:

Animal and history:

542 Living sheep of different ages, sexes and breeds were examined clinically in the field for investigation of the animals suffered from signs of respiratory problems [nasal discharge (serous – mucoïd - mucopurulent), lacrimation, and

coughing and abnormal lung sound by auscultation]. Those animals were reared in private farms at desert road and Sadat City. Others were reared in mobile flocks at Kafer Dawood village, Wadei lantron village and Elral village).

Sheep reared in the private farms were reared in properly ventilated at night and during the day they kept in wide yard with hygienic conditions. Good plane of nutrition in which the sheep fed daily on 1.5 kg concentrate hay or green bresseem also they were administered broad spectrum parasitic drug at regular interval as well as they vaccinated against various diseases. But those of mobile flocks were reared in bad hygienic conditions and poor nutrition. As they graze along the day on any area contain any feces at night they kept with other flocks in narrow and poorly ventilated moisten earthy ground. Dead sheep at farms and slaughtered at abattoirs subjected for postmortem examination, for collection of pathological lungs showing different degree of pneumonia (congestion, consolidation and hepatization).

13 Swiss mice about 18-22 gram weight were used for determination of virulence of the isolated *Pasteurella multocida*. These mice were obtained from the laboratory animal house at the Faculty of Veterinary Medicine Sidi Menoufia University.

Collection of samples and laboratory procedures:

Nasal swabs:

Sterile cotton swabs were used for collecting nasal swab samples from diseased (135) and apparently healthy sheep (50). (William et al., 1994)

Lung tissues:

105 lungs with various degree of pneumonia collected from freshly slaughtered animal at El-Bassatin, El-Moonib abattoirs and from died cases in the 20 normal lungs had been collected separately in sterile plastic bags and transported to laboratory on an ice tank within two hours of collection.

Bacteriological examination:

Primary isolation and purification:

Nasal swabs were incubated at 37°C for about 4 hours aerobically then the broth of the swab subcultured on blood agar, DAS medium, barker media and MacConkey's agar. As well as the surface of pneumonia tissues were sterilized using hot spatula and incision made using sterile then loopful of internal tissue were inoculated on the above mentioned media (Nadra, 1998).

Identification of the isolated strains:

Pure colony from each isolate was identified morphologically according to Gram staining reaction, shape, size and arrangement. Merchant and Packard, Blobel and Schliesser (1981) and Fingegold and Martin (1982)

The various members of family Enterbacteriaceae were typed biochemically according to Krieg and Holt (1984). *Pseudomonas* typed biochemically described by Quinn et al., (1994). Gram positive cocci included members of genera *Streptococcus* and *Staphylococcus*. were identified biochemically according to Queen et al., (1994).

The suspected isolates of *P. haemolytica* were typed biochemically into T and A types according to Buchanan and Gibbons (1974) and the pathogenicity test of *Pasteurella multocida* isolates were carried out according to (Amany, 1998).

Antibiotic sensitivity test of the isolates:-

This test performed by the disc diffusion method according to (Amany, 1998). The antibiotic discs were obtained from Oxoid were Amoxycillin 25µg, Erythromycin 10µg, Gentamycin 30µg, Peicillin G 10 units, Oxytetracyclin 100µg, Lincomycin 10µg, Enrofloxacin 10µg and Florofenicol 10µg.

Treatment trials:

Four protocols of treatment were performed:

The first protocol:

10% Enrofloxacin (Medtryl® Arabcomed) intramuscular injection 5mg / kg B.wt, for 5 days with 2.5% Diclofenac sodium (Diccloflam® Unipharma).1mg / kg. B.wt intramuscular injection for 2 days and AD3E+C (Advit C® Adwia) 2ml per animal injected intramuscularly for 5 days. This protocol applied upon 15 animals.

The second protocol:

Florfenicol (Nuflor® Schering-Pough) intramuscular injection 20mg / kg B.wt in two doses 48hrs apart with injection of Diclofenac sodium for 2 days and AD3E+C injected intramuscularly for 5 days. This protocol applied upon 15 animals.

The third protocol:

Oxytetracycline long acting (Terramycin L.A® Pfizer) 20mg / kg. B.wt. intramuscular injection in two doses 48hrs apart with Diclofenac sodium intramuscular injection for 2 days and AD3E+C injected intramuscularly for 5 days. This protocol applied upon 15 animals.

The forth protocol:

Only two injection of Amoxycillin long acting (Trioxy® Univet) 15mg / kg. B.wt. at 48 hrs intervals by intramuscular route with Diclofenac sodium intramuscular injection for 2 days and AD3E+C injected intramuscularly for 5 days. This protocol applied upon 15 animals.

Results

Description of the clinical signs and postmortem finding associated with respiratory affections of sheep in relation to bacterial isolate:

135 animals from 542 sheep showed various signs of respiratory disorder. From those 83 sheep showing sever respiratory signs and 52 sheep with mild signs.

Table (1) illustrates the clinical signs and postmortem examination in relation to main bacterial isolated from cases.

Main bacterial isolates	Clinical signs	Postmortem examination
Pasteurella haemolytica (alone or in combination with other bacteria)	Sever respiratory signs that include fever (ranged from 40°C to 42°C) with anorexia, depression. All these sheep showed bilateral mucopurulent nasal discharges and crusting around the nostrils and lacrymation fig. (1). Mouth breathing in most cases as well as cough was prominent and frequent and usually associated with expulsion of nasal discharges after coughing. Chest auscultations revealed exaggerated vesicular sound in some cases. In other cases moist rales with gargling sound was presented. In other cases no lung sound can be heard over the lung area.	Sever congestion all the lung with b purplish solid areas. the lung was he edematous with pete hemorrhage over lung, in the trachea at the heart Figure Other cases shc sever congestion hepatization of the ve part of diaphragn lobe. Figure (4). C cases showed s hyperemia and edem the l
Pasteurella multocida	Similar to the signs associated with pasteurella haemolytica	Congestion with pete hemorrhages and ma emphyse
Other bacterial agents : (Staph., Strept., Ecoli., Klebsiella., pseudomona s and salmonella) In single or mixed form.	Thick mucopurulent nasal discharges and seromucoid nasal discharges fig (2) with intermittent cough. The body temperature about 39.5°C, but chest auscultation in most of cases was dry rales and frictional sound.	Various degree congestion and gra discoloration of the with ecchym

Bacteriological findings:

Identification of bacterial isolates revealed the recovering of single and mixed isolates from the nasal swabs of diseased and lung tissues of slaughtered animals and from apparently healthy animal.

The following bacteria had been isolated from the respiratory tract of diseased and apparently healthy sheep (Pasteurella haemolytica, Pasteurella multocida)

E.coli, *Klebsiella pneumoniae*, *Salmonella*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Streptococcus pneumoniae*).

The percentage of infection and bacterial isolates were illustrated in table (2) and (3), respectively.

All the isolates of *Pasteurella haemolytica* belonged to *Pasteurella haemolytica* biotype A (recently called *Mannheimia haemolytica*) fig (5). All the isolates of *Pasteurella multocida* were pathogenic strain killed all inoculated mice within 24 hours after inoculation. Fig (6) showed *Pasteurella* bipolarity in stained blood smear of inoculated mice.

Table (2): The percentage of bacteria isolated from respiratory tract of apparently healthy and diseased sheep.

	Apparently healthy animals			Diseased animals		
	Number examined	+ ve for isolation No	%	Number examined	+ ve for isolation No	%
Nasal swab	50	35	70%	135	113	83.7%
Lung tissue	20	6	30%	105	75	71.42%
Total	70	41	58.57%	240	188	78.33%



Fig (1): Bilateral mucopurulent nasal discharge with crusting.



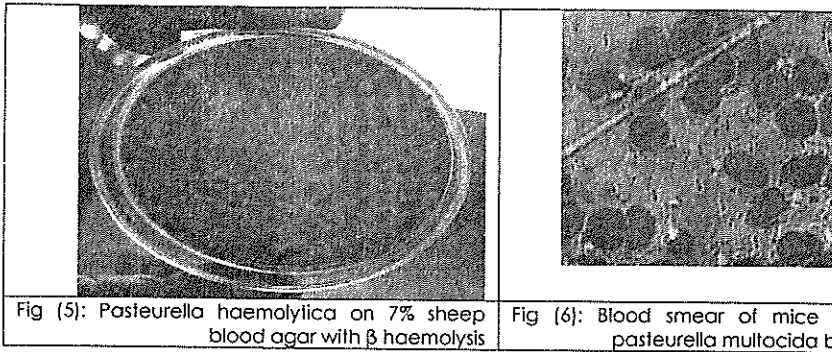
Fig (2): Thick mucopurulent nasal discharge



Fig (3): Lung congested with consolidation and pericarditis



Fig (4): Lung enlarged congested with emphysema and petechial haemorrhage



Discussion

In many countries respiratory diseases were the most serious sheep problem and an important cause of death and reduced productivity (Martin, 1996). Clinical examination revealed that the affected sheep suffering from fever (temperature above 40°C) with mucoid to mucopurulent nasal discharges and crusting around the nostrils with depression in some cases. Chest auscultation was the presence of abnormal lung sounds as moist rales, exaggerated vesicular sounds as well as bubbling sound also were heard in some cases. In other cases no lung sound could be detected over the lung area but the heart sound is clear from the right chest side. From these signs the affected sheep were suspected to be suffering from bronchopneumonia. These signs were recorded by Yousif, 1981; Elyas, 1993; Sadiek et al., 1993; Abdel-Salam and Abd-El-Moneim, 1994; Mary and David, 1994; Martin, 1996 and Zaitoun, 2001, they described the clinical findings in sheep suffering from acute bacterial pneumonia.

The results of clinical examination are also supported by the records of Donachie, 2000; Radostitis, et al., 2000 and Mona et al., 2005 as they described mild clinical disease with seromucoid and thick tenacious nasal discharges and intermittent cough. Chest auscultation reveals the presence of dry rales and friction sound.

Table (2) illustrated the incidence of bacterial isolation from nasal swabs was 70% and 30% from apparently normal lung. These figures were nearly similar to that obtained by Faten, 2001 but less than that obtained by Nadra, 1998. The bacterial isolates recovered from apparently healthy sheep were *Pasteurella haemolytica*, *E. coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Staphylococcus epidermidis*. Mixed isolates of *Pasteurella haemolytica* with *E. coli* and *E. coli* with *Staphylococcus aureus* and with *Staphylococcus epidermidis* were also recovered. This result agreed with the finding of El-Shorbagy and Abd-El-Ghani, 1974 and Nadra, 1998. On the other hand Faten, 2001 isolated *Pasteurella multocida* and *Corynebacterium pseudotuberculosis* from apparently healthy sheep. The recovery of the different bacterial isolates from the nasal swabs of apparently healthy sheep referred to the presence of carrier animals within the herd representing a source of infection to other sheep in the herd. While the isolation of bacterial agents from apparently normal lung

may be indicating subclinical infection or from contamination of lung tissue during the bad handling of carcass in the abattoirs. It well known that about 40% of healthy animals carry *Pasteurella* species in the upper respiratory tract that under the stress factors causing respiratory disease (Radostitis, 2000 and The Merck Veterinary Manual, 2006)

Bacteriological examination of the nasal swabs of diseased sheep as well as pneumonic lung tissues revealed the presence of many bacterial species as a single isolates or in mixed forms. This agree with many authors as Alley and Clark, 1980; Nadra, 1998; Faten, 2001; Zaitoun, 2001 and Mona et al., 2005. The following bacterial isolates had been isolated from nasal swabs of diseased sheep and pneumonic lung tissue; *Pasteurella haemolytica*, *Pasteurella multocida*, *E.coli*, *Klebsiella pneumonia*, *Salmonella*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Streptococcus pneumoniae*. These bacterial isolates also isolated by Sambyal et al., 1980; Baysal and Guler, 1992; Queen et al., 1994; Nadra, 1998 and Mona et al., 2005. The percentage of bacterial isolation from the diseased animals was 78.33% which includes 113 isolates out of 135 examined nasal swab of diseased sheep (83.7%) and 75 out of 105 pneumonic lung tissue examined (71.42%). This was similar to the incidence of bacterial isolation by Kaya and Erganis, 1991 and Mona et al., 2005. But less than the incidence of bacterial isolation by Elsherif and Abdel-Ghani 1974, Nadra, 1998, and Faten, 2001.

Pasteurella haemolytica was the more commonly isolated bacteria from diseased sheep either from nasal swabs or pneumonic lung tissues. Table (3) presented the percent of isolation of *Pasteurella haemolytica* were 26.54% and 26.66% from the nasal swabs or pneumonic lung tissues, respectively. On the other hand *Pasteurella multocida* was unusually isolated from the diseased sheep. Only the percent of its isolation were 3.54% and 12% from nasal swabs and pneumonic lung tissues, respectively. These result were in accordance with Elsherif and Abdel-Ghani, 1974; Davis, 1985; Younan et al., 1988; Kaya and Erganis, 1991; Baysal and Gular, 1992; Queen et al., 1994; Boulijihad and Leipold, 1995; Mohamed, 1996; Nadra, 1998; Faten, 2001; Gelagay et al., 2004 and Mona et al., 2005. Hancock et al., 1991 recorded that *Pasteurella multocida* was the common isolated bacteria from pneumonic sheep.

Pasteurella haemolytica isolated in combination with *E.coli* at percent of 9.04% which includes 10 isolates from nasal swab (8.85%) and 7 isolates from pneumonic lung tissues (9.33%). This agreed with Nadra, 1998. Mona et al., 2005 isolated also *Pasteurella multocida* in combination with *E.coli* from nasal swab and lung tissues of diseased sheep. *Pasteurella haemolytica* was also isolated in combination with gram positive bacteria mainly *Staphylococcus aureus* (5.31%) which includes 7 isolates from nasal swab (6.19%) and 3 isolates from pneumonic lung tissues (4%). Nadra, 1998 isolated *Pasteurella haemolytica* in combination with gram positive bacteria which in his case was *Streptococcus pneumoniae* from the nasal swabs at percent of 9%. Biotyping of *Pasteurella haemolytica* isolates in this study revealed that 100% of the isolates belong to *Pasteurella haemolytica* biotype A (*Mannheimia haemolytica*). This agreed with the result obtained by Blanco-viera, et al., 1995. The results obtained by Guler et al., 1996 and Mohamed, 1996 indicated the

isolation of both biotype A and T with also untypable strains. Dunbar et al., 1990 isolated both biotypes from tonsillar biopsies rather than from the nasal swabs. Pathogenicity test of *Pasteurella multocida* revealed that all *Pasteurella multocida* isolates were pathogenic to the mice. These results were in accordance with that recorded by Amany, 1998. On the other hand Zincir, 2004, illustrated that only 86.7% of the isolated *Pasteurella multocida* from sheep suffering from respiratory disease were positive in mice pathogenically test.

Table (3) illustrated the percent of isolation of different members of the family Enterobacteriaceae. *E. coli* was the common members of this family frequently isolated from both nasal swab as well as from pneumonic lung tissues. The total percent of isolation was 12.23% that include 17 isolates from nasal swabs (15.04%) and 6 isolates from pneumonic lung tissues (8%). These results agreed with El-sherif and Abdel-Ghani, 1974; Faten, 2001 and with Mona et al., 2005. Sambyal et al., 1980 recorded a percentage of its isolation of 27.7%. Elyas, 1993; Nadra, 1998 and Zincir, 2004 recorded a percentage of isolation of 3%, 7.7% and 2.8%, respectively. The differences between records were mainly due to the geographical distribution at which the investigator was adopted. *E. coli* isolated in combination with *Staphylococcus aureus* from both nasal swabs and pneumonic lung tissues at percent of 10.63% and 8% respectively. These results agreed with Zincir, 2004 and Mona et al., 2005. *Klebsiella pneumoniae* isolated from diseased sheep at percent of 7.96% which includes 9 isolates from nasal swabs (7.96%) and 6 isolates from pneumonic lung tissues (8%). This agreed with Ikede 1978; Kaya Erganis, 1991; Gameel et al., 1991; Elyas 1993; Queen et al., 1994; Nadeem, 1998; Faten, 2001 and Mona, et al., 2005. Sambyal et al., 1980 recorded a percentage of its isolation of (17%). *Klebsiella pneumoniae* had been isolated in combination with *Pseudomonas aeruginosa* from nasal discharge of diseased sheep. *Salmonella* had been isolated from the nasal swabs of diseased sheep at percent of 3.54%. These results agreed with that obtained by Mona et al., 2005. Table (3) presented the isolation of *Pseudomonas aeruginosa* from nasal swabs and pneumonic lung tissues. The total percentage of its isolation was 5.85% that include 8 isolates from nasal swabs (7.079%) and 3 cases from pneumonic lung tissues (4%). These results agreed with that obtained by El-sherif and Abdel-Ghani, 1974; Nadra, 1998 and Mona et al., 2005. On the other hand Sambyal et al., 1980; Baysal and Guler, 1992 and Zincir, 2004., recorded a lower percentage of isolation of *Pseudomonas aeruginosa*.

Staphylococcus aureus had been isolated at percent of 10.63% that include 12 isolates from nasal swabs (10.61%) and 8 isolates from pneumonic lung tissues (10.66%). That result agreed with Baysal and Gular, 1992; Sukhon, 1995; Nadra, 1998 and Mona et al., 2005. On the other hand El-sherif and Abdel-Ghani, 1974; Sambyal et al., 1980 and Elyas, 1993. recorded a percentage of isolation of 6.2%, 3.1% and 6%, respectively. Zincir, 2004, recorded a percentage of isolation of 17.7%. *Streptococcus pneumoniae* isolated at percent of 6.38% that include 5 isolates from nasal swabs (4.44%) and 7 isolates from pneumonic lung tissues (9.33%). These results agreed with El-sherif and Abdel-Ghani, 1974; Sambyal et al., 1980, Elyas, 1993; Nadeem, 1998 and Mona et al., 2005. On the other hand Zincir, 2004., recorded a lower

percentage of isolation. The antibiogram of isolated bacteria shown that most isolates were sensitive to Flurofenicol, Enrofloxacin, Amoxycillin and Gentamycin. While moderate sensitivity showed against Oxytetracycline, Penicillin and Erythromycin. But most of them were resistant to Lincomycin. This was similar to the antibiogram result of Zaitoun, 2001 and Mona et al., 2005. While it differed with Faten, 2001 who reported that most bacterial isolates recovered from pneumonic sheep are highly sensitive to Oxytetracycline and Erythromycin but she did not test these bacteria against Enrofloxacin, Amoxycillin.

The different treatment trials applied on the diseased animals using different antibacterial agent indicating that the rapid and complete recovery result was from florofenicol administration. The use of enrofloxacin also showed good result that agreed with Zaitoun, 2001, and similar to the findings obtained by Cusack et al., 2003. Who found that flurofenicol was used to effectively reducing replase and case fatality rates. Also he found that enrofloxacin had been used to effectively respiratory diseases in USA. and Europe. Also agreed with Ronald et al., 2004 who advise the use of aggressive antimicrobial therapy using one or combination of tilmicosin, florofenicol, cetiofur or enrofloxacin. Long acting amoxycillin had been given good results in treatment of infected cases. The unstayfactory results were by using of oxytetracycline this might be due to development of bacterial resistance against /the oxytetracycline. Which disagreed with Nadra, 1998; Donachie, 2000; Faten, 2001. They recorded that the Oxytetracycline gives good result in treatment of pneumonic cases.

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الملخص العربي

من خلال هذه الدراسة تبين أن الأصابات التنفسية في الأغنام توجد في صورتين . الأولى تكون في صورة شديدة مع أعراض حادة و الأخرى في صورة مزمنة وأقل حدة مع كحة مستمرة و ضعف عام. كما تم عزل البكتريا المسببة للإصابة في صورة منفردة ومختلطة. مثل البستريلا هيموليتيكا والبستريلا ملتوسيدا و ايشيريشيا القولون و الكلبسيلا الألتهاب الرئوي و السلمونيلا و بكتريا الصديد الأخضر وبكتريا العنقود الذهبي وبكتريا المكروب السبحي الرئوي كم تم ربط التغيرات الباثولوجية مع البكتريا المعزولة. اوضحت نتيجة أختبار الحساسية للبكتريا المعزولة أن كل البكتريا المعزولة حساسة لعقار الفلورفينيكول و الأنتروفلوكساسين وأقل حساسية لكل من الجنتاميسين والأموكسللين و مقاومة لكل من اللنكوميسين و الأوكسي تتراسيكلين والبنسلين. ومن خلال تجارب العلاج كان كل من الفلورفينيكول و الأنتروفلوكساسين هما الأكثر سرعة في معدل الشفاء وتحسن الحالات.

Antibiogram of the isolated bacteria:

The antibiogram of isolated bacteria shown that most isolates were sensitive to Fluorfenicol, Enrofloxacin, Amoxicillin and Gentamycin, as shown in table (4).

Table (4) showed the result of antibiogram test.

	Enrofloxacin	Amoxicillin	Erythromycin	Gentamycin	Penicillin-G	Oxytetracycline	Fluorfenicol	Lincomycin
1-Pasteurella haemolytica	+++	++	+	++	+	++	+++	+++
2-Pasteurella multocida	+++	++	-	++	+	+	+++	+++
3-E. Coli	+++	++	-	+	-	+++	+++	+
4-Klebsiella pneumoniae	++	-	-	++	-	+	++	-
5-Salmonella	++	+	+	++	-	++	++	-
6-pseudomonas aeruginosa	+++	+	+	++	-	-	++	-
7-Staphylococcus aureus	++	+++	+	-	++	+	+	-
8-Streptococcus pneumoniae	++	++	+	++	+	-	+	-

Treatment trials:

Treatment protocol	Cases progress
First	10 cases completely recovered after 3 days. 5 cases recovered after 5 th day of treatment
Second	All 15 cases completely recovered after 1 st dose.
Third	6 cases recovered completely after 3 rd day. 9 cases recovered after 5 th day of treatment
fourth	12 cases recovered after 3 days.

Table (3): The percentage of each bacterial isolate from respiratory tract of apparently healthy and diseased sheep

Bacterial isolates	Apparently healthy				diseased					
	Total		Lung tissue %		Total		Lung tissue %			
	No	%	Nasal swab No	%	No	%	Nasal swab No	%		
A- Single isolates:										
1-Pasteurella haemolytica	5	12.2%	5	14.3%	-	-	30	26.54%	20	26.66%
2-Pasteurella multocida	-	-	-	-	-	-	4	3.54%	9	12.00%
3-E.Coli	6	14.6%	4	11.4%	2	33.33%	17	15.04%	6	8.00%
4-Klebsiella pneumoniae	-	-	-	-	-	-	15	7.97%	6	8.00%
5-Salmonella	-	-	-	-	-	-	4	2.13%	4	3.54%
6-pseudomonas aeruginosa	2	5%	2	5.7%	-	-	11	5.85%	8	7.079%
7-Staphylococcus aureus	7	17.1%	6	17.4%	1	16.60%	20	10.63%	12	10.61%
8-Streptococcus pneumoniae	-	-	-	-	-	-	12	6.38%	5	4.42%
7-Staphylococcus epidermidis.	10	24.4%	8	23%	2	33.33%	-	-	-	-
B-Mixed isolates:										
1-P.haemolytica+E.Coli	2	5%	2	5.7%	-	-	17	9.04%	10	8.85%
2-P.haemolytica+Staph.aureus	-	-	-	-	-	-	10	5.31%	7	6.19%
3-E.Coli+Staph.aureus	2	5%	2	5.7%	-	-	12	6.38%	6	5.4%
4-Klebsiella pneumoniae+Pseudomonas aeruginosa	-	-	-	-	-	-	1	0.53%	1	0.88%