

EVALUATION OF SELECTED *Azospirillum* SP. ISOLATES FOR IAA PRODUCTION AND THEIR POTENTIAL IMPACT ON IMPROVING GROWTH, YIELD AND FRUIT QUALITY OF 'ANNA' APPLE TREES

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ABSTRACT

Foliar spray with the diluted PGPR (plant growth promoting rhizobacteria) cultures had utmost importance due to its content and release of stimulants, nutrients, antibiotics, biocides and siderophores. Potentiality of these microorganisms, in plant rhizosphere, in activation and improving of plant growth as well as increasing plant tolerance to different plant biotic and abiotic stresses have been proved. However, microbial types and even microbial strains varied for their potentiality to adapt, inhabit and release of stimulants and phytochemicals.

A number of *Azospirillum* sp. isolates were isolated from phyllosphere of apple, orange, lemon, mango and guava trees, grown at El-Bostan area, Egypt. These isolates were used in spray of apple trees with the dose of 20 and 40 L/feddan compared to water spray control. Foliar spray of apple trees with all *Azospirillum* isolates notably induced the plant growth and increased fruit yield, but did not significantly affect fruit quality. However, the used isolates largely varied in their efficiency and potentiality. A⁷ isolate was the superior followed by A⁹ which attained the highest increases in shoot length, shoot diameter, leaf area, leaf dry weight leaf content of chlorophyll a, b and total chlorophyll as well as fruit yield over those of control treatment. But, the quality parameters, i. e., SSC (soluble solids content), acidity, firmness and color did not show consistent significant variations. The treatments of spray with *Azospirillum* isolates attained high increase in net return (L E/fed), the spray with the treatments of A⁹D (isolated from mango phyllosphere and used with the rate of 40 L/fed.) and A⁹R, which isolated from mango phyllosphere and used with the rate of 20 L / fed.) were the superior treatments which achieved increases in net return over that of control (water spray) by 6120 and 6200 L E/fed respectively, followed by A⁷R (isolated from phyllosphere of apple and applied with 20 L/fed.) which gave 5990 L E/fed. It is noted that the microbial culture dose of 40 L/fed had no significant differences than the application of 20 L/fed. Therefore, we recommend the spray with an efficient *Azospirillum* isolate like A⁷ or A⁹ with the dose of 20 L/fed, and it is also of importance to conduct studies on isolation and evaluation of these microorganisms to select the most efficient strains for use, as inoculants, in spray of apple trees.

INTRODUCTION

The Egyptian economy needs to be improved; this requires us to find out practical and applicable solutions for improving plant yield. The foliar spray with PGPR bacteria had been proved efficiency for enhancing plant growth and yield of different crops (Esitken *et al.*, 2009; Sekar and Kandavel, 2010 and Ryu *et al.*, 2011). Saharan and Nehra (2011) attributed the enhancement effect of PGPR to their direct effect in releasing plant growth hormones, nitrogen fixation, increase of plant potentiality to absorb nutrient elements and release of siderophores, which chelate Fe and making it

available for plant use. Moreover, the PGPR containing ACC (1-Aminocyclopropane-1-Carboxylate) deaminase are present in various soils and offer promise as a bacterial inoculum for improvement of plant growth, particularly under unfavorable environmental conditions such as flooding, heavy metals, phytopathogens, drought and high salt (Belimov *et al.*, 2001). Also, production of biotoxins and antibiotics by these PGPR gave the plant high potentiality to resist pathogens (Anith and Momol, 2004), in addition spray with PGPR filling phyllosphere area at the expense of harmful microbes.

It was found that the potentiality of PGPR types in inducing plant growth varied from type to another and from strain to another in the same species (Dursun *et al.*, 2010). The ability of the IAA production, antibiotic synthesis and N₂-fixation are variable (Fernando *et al.*, 2005; Aslantas *et al.*, 2007 and Saharan and Nehra, 2011). PGPR was found also to modify the plant hormones status (Dodd *et al.*, 2010). Therefore, it is of importance to select high effective strains had potential influence in increasing plant growth which, consequently, will reflect on the agricultural economics and national income.

Therefore, the present investigation aims to study the potentiality of different *Azospirillum* isolates, which isolated from the phyllosphere of different fruit trees on producing IAA and enhancing apple tree growth, productivity and fruit quality.

MATERIALS AND METHODS

Materials:

Microbial media:

Azospirillum: Semi solid Döbereiner medium (Döbereiner *et al.*, 1976) contained (g/liter of distilled water): Malic acid, 0.5; KH₂PO₄, 0.5; K₂HPO₄, 0.5; MgSO₄. 7H₂O, 0.5; NaCl, 0.5; CaCl₂. 2H₂O, 0.5; FeCl₃. 6H₂O, 0.5; Na₂MoO₄. 2H₂O, 0.05; agar, 1.5.

This study has been carried out on eight years old "Anna" apple trees budded on Malus rootstock during 2009 and 2010. Trees were grown at Elbostan region of Elbehira Governorate, where drip fertigation system was applied and soil texture analysis was shown in Table 1.

Table 1: Some chemical and physical analysis characteristics of the experimental soil

Sand %	Silt %	Clay %	Texture	O.M %	pH	EC (ds m ⁻¹)			
80.2	18.0	11.3	Sandy clay loam	0.62	8.1	1.19			
Cations (mg./L)			Anions (mg./L)			Macro-nutrient (mg/kg)			
Na ⁺	Ca ⁺⁺	Mg ⁺⁺	CO ₃ ⁻⁻	HCO ₃ ⁻	Cl ⁻	SO ₄ ⁻⁻	N	P	K
0.89	10.42	3.91	-	1.37	12.41	6.44	139	7	86

Methods:

The *Azospirillum* bacteria were isolated from phyllosphere of different fruit trees (lemon, guava, apple, orange and mango). The liquid culture of the different isolates was used for spray the experimental apple trees and the densities ranged between 1 to 10⁸ X 10⁶ cfu/ml culture. Treatments were

arranged in a random order on the selected trees. Single tree plot with 3 replicates for each treatment was arranged in random complete blocks design.

Bacterial strains were tested for their capability to produce indole acetic acid (IAA) (Bric *et al.*, 1991)

All trees were subjected with common regional horticultural practices, while treatments were applied as follows in Table (2):

Treatment	Description
A 1 R (Lemon)	Spray with <i>Azospirillum</i> isolated from lemon phyllosphere, 20 L / feddan.
A 1 D (Lemo)	Spray with <i>Azospirillum</i> isolated from lemon phyllosphere, 10 L / feddan.
A 2 R (Guava)	Spray with <i>Azospirillum</i> isolated from guava phyllosphere, 20 L / feddan.
A 2 D (Guava)	Spray with <i>Azospirillum</i> isolated from guava phyllosphere, 10 L / feddan.
A 3 R (apple)	Spray with <i>Azospirillum</i> isolated from apple phyllosphere, 20 L / feddan.
A 3 D (apple)	Spray with <i>Azospirillum</i> isolated from apple phyllosphere, 10 L / feddan.
A 4 R (Orange)	Spray with <i>Azospirillum</i> isolated from orange phyllosphere, 20 L / feddan.
A 4 D (Orange)	Spray with <i>Azospirillum</i> isolated from orange phyllosphere, 10 L / feddan.
A 5 R (Mango)	Spray with <i>Azospirillum</i> isolated from mango phyllosphere, 20 L / feddan.
A 5 D (Mango)	Spray with <i>Azospirillum</i> isolated from mango phyllosphere, 10 L / feddan.
Control	Spray with water.

The microbial inoculants were prepared in Soil Bacteriology Laboratory of Sakha Agricultural Research Station, ARC. Bacterial suspension was diluted by mixing 20 L or 10 L of bacterial stocks with 100 L of water per feddan.

Three branches, five years old, in different directions on each tree were selected and labeled to estimate growth parameters. All current shoots developed on these branches were measured to get shoot length (cm). Li-Core-3100 Areameter was used to measure detached leaves of nine shoots (three shoots per branch) to get area per leaf (cm²). Leaves were dried at 70°C and weighed to get dry weight (mg) and then specific leaf weight (SLW) was calculated as (mg cm⁻²).

Spectrophotometer was used to estimate chlorophyll a and chlorophyll b, which extracted from fresh leaves with di-methyl formamide (DMF) as described by Rami and Porath (1980). The concentration of chlorophyll a and chlorophyll b and its total value were calculated by Rami's formulas as (µg / ml) (Rami, 1982). The results were presented as (mg.cm⁻²).

Fruits were picked at maturity stage, weighed and counted. Fruit pulp texture (firmness) was recorded by using Lfra texture analyzer instrument. The results were expressed as a resistance force of the fruit to the penetrating tester (g/cm²) according to Harold (1980). Fruit skin color measurements (a*, b*, L* & H°) were determined using Minolta colorimeter (Minolta Co. Ltd., Japan). The instrument estimated skin color of fruits with color metric CIE Lab method where L* measure lightness scale readings and the two coordinates a* and b* included. Positive values of a* is a measure of redness and becomes greenish measure when values changed into negative, while b* of yellowness and blueness (- b*) on the Hue circle. The Hue angle [H° = arc tan (b*/a*)] describe the relative amounts of redness and yellowness where point at 0°/360° is defined for red/magenta, 90° yellow, 180° for green and 270° for blue color (McGuire, 1992 and Voss, 1992).

Soluble solids content (SSC) was determined by using a hand refractometer and total acidity percentage was estimated in filtered juice according to A.O.A.C. (1990).

Statistical analysis:

Data obtained were subjected to the analysis of variance and treatment means were compared using the L.S.D. methods according to Steel and Torrie (1980).

RESULTS AND DISCUSSION

Carefully considering readings of Table 3, spraying with different *Azospirillum* isolates that have been isolated from phyllosphere of different tree species i.e. apple, orange, lemon, mango and guava, the foliar spray of apple trees with these isolates positively affected shoot length with varied degrees, and this trend was consistent throughout both seasons of study, but the differences than water spray control treatment were significant at the second season, only. The spray with A²D isolate gave the highest difference throughout both studied seasons.

Table 3: Response of vegetative growth of "Anna" apple trees to spray with phyllosphere *Azospirillum* isolates.

Treatments	Shoot length (cm)		Shoot diameter (cm)		Leaf dry weight (g)		Leaf area (cm ²)		SLW (mg cm ⁻²)	
	2009	2010	2009	2010	2009	2010	2009	2010	2009	2010
A ¹ R (Lemon)	07.8 c	04.7 a	1.6 a	1.0 def	0.36 cde	0.33 c-g	34.0 b	32.0 a	10.6 ab	10.1 c
A ¹ D (Lemon)	09.0 bc	00.7 a	1.7 a	1.0 cd	0.38 bcd	0.30 b-e	36.6 ab	34.4 a	10.2 b	10.1 bc
A ² R (Guava)	06.8 c	04.1 a	1.6 a	1.4 f	0.33 efg	0.30 efg	26.0 cd	24.3 bc	12.6 a	12.2 abc
A ² D (Guava)	08.3 c	04.0 a	1.6 a	1.0 def	0.30 def	0.32 d-g	27.7 cd	20.3 bc	12.0 ab	12.6 abc
A ³ R (apple)	17.4 a	13.2 a	1.7 a	1.6 bc	0.41 ab	0.38 ab	38.4 ab	30.9 a	10.6 ab	10.6 abc
A ³ D (apple)	16.9 a	14.0 a	1.7 a	1.6 ab	0.42 a	0.39 a	39.3 a	36.8 a	10.7 ab	10.2 bc
A ⁴ R (Orange)	07.0 c	00.1 a	1.6 a	1.0 def	0.36 cde	0.33 e-g	29.7 c	20.9 bc	12.8 a	12.7 ab
A ⁴ D (Orange)	07.8 c	04.9 a	1.6 a	1.0 de	0.36 cde	0.34 b-f	28.7 cd	27.4 b	12.4 ab	12.2 abc
A ⁵ R (Mango)	11.9 bc	09.1 a	1.7 a	1.6 bc	0.39 a-d	0.36 a-d	36.4 ab	33.9 a	10.6 ab	10.0 abc
A ⁵ D (Mango)	14.9 ab	11.0 a	1.7 a	1.7 a	0.39 bc	0.37 abc	37.4 ab	34.9 a	10.0 ab	10.0 abc
C (water spray)	06.4 c	04.0 b	1.0 b	1.4 b	0.30 g	0.28 d	24.4 d	22.3 g	12.0 ab	12.0 a-d

AR: *Azospirillum* spray with 10 L / feddan; AD: *Azospirillum* spray with 40 L / feddan, SLW : Specific leaf weight.

Means with different letters within the same column differ significantly at P < 0.05.

Data of shoot diameter pursued the same approach of shoot length which similarly increased due to the foliar application of *Azospirillum* isolates. The increases over control mostly were significant at the second season. Likewise, the spray treatments attained big and consistent increments in leaf dry weight and leaf area at both seasons, the differences than control, mostly, were significant. It is also noted that the application of A^r isolate achieved the highest values, i. e., 0.42 and 0.39 g/leaf, and 39.3 and 36.8 cm² for leaf area, respectively. While, the spray treatments did not affect the SLW values, whereas, all the differences than control, at both seasons, were not significant.

The spray of apple trees with the different bacterial strains had an effective role in increasing shoot length and diameter, leaf area and leaf dry weight compared to control. These results are in harmony with those of Eissa *et al.* (2007) who indicated that the spray of pear trees with *Saccharomyces cerevisia* had a stimulated effect on tree growth. While, Aslantas *et al.* (2007) reported that applications with different types of PGPR lead to significant increases in shoot length and diameter and they attributed this effect to the potentiality of the bacteria for releasing cytokinins and IAA. They found also that plant growth responses were variable and dependent on bacterial strain. These results agreed also with the results of the present study which revealed that the different isolates of *Azospirillum* had varied stimulation potentiality to the plant growth and the best isolate was that isolated from phyllosphere of apple (A^r).

The readings of Table 4 showed that foliar spray with *Azospirillum* isolates increased leaf chlorophyll content. Whereas, chlorophyll a showed high and significant increases over control, the influence at the first season higher than those of the second season. Similarly, the spray treatments attained remarkable increases in chlorophyll b content, the increases were obvious at first season and the most differences than control were significant. The total chlorophyll content exhibited the same trend.

Table 4: Response of chlorophyll content of "Anna" apple leaves to spray with phyllosphere *Azospirillum* isolates.

Treatments	Chlorophyll a (mg cm ⁻²)		Chlorophyll b (mg cm ⁻²)		Total chlorophyll (mg cm ⁻²)	
	2009	2010	2009	2010	2009	2010
A ^r R (Lemon)	4.89 a	4.93 ab	2.40 a	2.30 cde	7.29 a	7.07 bc
A ^r D (Lemon)	4.07 a	4.78 bc	2.00 a	2.30 bcd	6.07 b	7.08 c
A ^r R (Guava)	4.36 a	4.69 bc	2.20 a	2.08 ef	6.56 d	6.70 e
A ^r D (Guava)	4.30 a	4.12 ef	2.20 a	2.06 ef	6.50 d	6.18 e
A ^r R (apple)	4.90 a	4.18 ef	2.08 a	2.08 ab	6.98 a	6.26 ab
A ^r D (apple)	4.90 a	4.78 ab	2.78 a	2.61 a	7.68 a	7.39 abc
A ^r R (Orange)	4.60 a	4.72 abc	2.37 a	2.21 def	6.97 c	6.99 d
A ^r D (Orange)	4.66 a	4.38 de	2.37 a	2.28 def	7.03 c	6.66 d
A ^r R (Mango)	0.12 a	4.21 ef	2.62 a	2.47 abc	2.74 a	6.68 a
A ^r D (Mango)	0.10 a	4.96 a	2.62 a	2.48 abc	2.72 a	7.44 a
C (water spray)	2.96 b	4.09 f	1.06 b	2.14 def	4.02 d	6.23 e

AR: *Azospirillum* spray with 20 L / feddan; AD: *Azospirillum* spray with 10 L / feddan.
Means with different letters within the same column differ significantly at P < 0.05.

Whereas, leaf total chlorophyll content increased with high and significant values due to spray with the different *Azospirillum* isolates. The application of isolates used in treatments A^z and A^y attained the highest values of total chlorophyll content.

Spray of apple trees with *Azospirillum* isolates significantly increased the contents of chlorophyll a, b, and total chlorophyll of leaves, these results are harmony with those of Eissa (2003) who found that spray of apricot with dry yeast extract increased chlorophyll contents of leaves. This may attributed to the release of beneficial compounds as polyamines (Babalola, 2010), which were found to increase chlorophyll a, b and carotenoids (Taha and Eid, 2011).

The beneficial effect of spray with *Azospirillum* isolates reflected, also, on the fruit yield (Table 2) whereas, number of fruits/tree notably increased. The fruits weight (kg/tree), at the both seasons was increased too, with high consistent and significant values compared to control treatment. The highest productivity (40.93 and 43.23 kg/tree) were recorded for A^zD treatment and 40.30 and 41.96 kg/tree for A^zR treatment followed by A^yD and A^yR treatments, which attained 40.20 and 41.93 kg/tree, and 44.43 and 41.06 kg/tree at seasons 2009 and 2010, respectively. The results of IAA values of the studied *Azospirillum* isolates followed the same trend of fruit yield, whereas, the highest IAA values were achieved by application with A^z treatment (3.40 µg/ml of culture) followed by A^x and A^y which exhibited, to some extent, similar values i. e. 2.80 and 2.76 µg/ml respectively. The treatment of A^z was exhibited the lowest IAA release that estimated 1.81 µg/ml of culture.

The present study showed that spray with all *Azospirillum* isolates significantly increased yield of apple fruits per tree, and the spray with the isolate A^z, which isolated from apple leaves, were absolutely the best followed by the isolate which isolated from mango leaves. These results are completely agreed with those of Esitken *et al.* (2004) who reported that spray of apricot with *Bacillus* OSU-142 increased fruit yield. Similarly, Eissa (2003) reported that the spray with EM resulted in an increase in number and weight of "Kelsey" plum fruits/tree. Also, Eissa *et al.* (2007) indicated that the spray of pear trees with *Saccharomyces cerevecia* had a stimulated effect and increased number and weight of fruits. Martinez-Viveros *et al.* (2010) summarized the mechanisms of PGPR action on plant growth as follow: the plant growth stimulation by PGPR is the net result of multiple mechanisms of action:

- 1- Microorganisms having mechanisms that facilitate nutrient uptake or increase nutrient availability as fix of nitrogen or solubilizing phosphates and mineralize organic compounds.
- 2- Production of phytohormones is now considered to be one of the most important mechanisms by which many rhizobacteria enhance plant growth, like IAA.
- 3- Regulate plant ethylene levels, the high accumulation of ethylene leads to poor roots growth leads a diminished ability to acquire water and nutrients.

ε- Can provide biocontrol of diseases or insect pests (biopesticides) via production of antibiotics, siderophores, HCN, hydrolytic enzymes (chinases, proteases, lipases... etc.).

The present study (Table 6) indicated that the different isolates released IAA differently and A^r isolate had the highest level of IAA production and also tree fruit yield, followed by isolate A^y.

Table 6: Response of "Anna" apple yield to spray with *Azospirillum* isolates and the concentration of IAA in culture

Treatments	Fruit weight / tree (kg)		Fruit number / tree		IAA (µg/ml)
	2009	2010	2009	2010	
A ^r R (Lemon)	44.00 ab	40.72 abc	34.0 a-d	312 ab	2.80
A ⁱ D (Lemon)	44.06 ab	40.96 abc	312 def	314 ab	
A ^r R (Guava)	40.60 b	37.36 c	298 def	260 de	1.98
A ^y D (Guava)	41.36 ab	37.80 bc	301 def	270 de	
A ^r R (apple)	40.30 a	41.96 ab	367 ab	330 a	3.40
A ^y D (apple)	40.92 a	42.22 a	374 a	336 a	
A ⁱ R (Orange)	43.10 ab	39.42 abc	323 c-f	200 def	1.81
A ⁱ D (Orange)	43.46 ab	40.20 abc	327 b-e	286 de	
A ^y R (Mango)	44.42 ab	41.06 abc	368 ab	270 de	2.76
A ^y D (Mango)	40.20 a	41.92 ab	362 abc	301 de	
C (water spray)	33.72 c	32.12 d	292 ef	291 de	-

AR: *Azospirillum* spray with 20 L / feddan; AD: *Azospirillum* spray with 40 L / feddan. Means with different letters within the same column differ significantly at P < 0.05.

Data presented in Table 7 showed the effect of foliar spray with *Azospirillum* isolates on some determinations of apple fruit quality. The different treatments, in general, did not exhibit significant influence on juice soluble solids content (SSC) percentage and firmness. While, the effect of different isolates on fruit acidity percentage was varied. In spite of incidence of decrease of fruit acidity due to the spray with *Azospirillum*, Aⁱ isolates attained significant increase, whilst not reached significance in case of A^r isolate.

Table 7: Response of "Anna" apple fruit quality to spray with phyllosphere *Azospirillum* isolates

Treatments	SSC %		Acidity %		Firmness (g/cm ³)		Color	
	2009	2010	2009	2010	2009	2010	2009	2010
A ^r R (Lemon)	12.30 d	12.63 d	0.90 a	0.86 a	227.0 abc	274.3 a	89.16 ab	82.80 ab
A ⁱ D (Lemon)	12.43 cd	12.76 cd	0.89 a	0.86 a	236.0 abc	269.3 a	88.20 abc	83.90 ab
A ^y R (Guava)	12.43 cd	12.80 bcd	0.88 a	0.80 abc	220.3 bc	234.3 bc	92.10 ca	86.02 a
A ^y D (Guava)	12.03 bcd	12.73 d	0.87 ab	0.84 abc	236.0 abc	241.3 abc	90.17 ab	86.07 a
A ^r R (apple)	12.80 abc	13.16 abc	0.81 cd	0.78 b-e	243.3 abc	206.1 ab	83.17 bcd	78.20 bc
A ^y D (apple)	12.80 abc	13.16 abc	0.82 bc	0.79 bcd	246.3 abc	208.2 ab	82.18 b-e	77.81 bc
A ⁱ R (Orange)	12.90 ab	13.20 ab	0.74 e	0.71 ef	243.3 abc	290.3 ab	80.23 c-f	71.07 cd
A ⁱ D (Orange)	12.92 ab	13.26 a	0.70 cde	0.71 def	246.1 abc	208.1 ab	73.9 ef	69.68 d
A ^y R (Mango)	12.92 ab	13.23 a	0.73 e	0.69 f	262.0 a	274.3 a	71.67 f	66.70 d
A ^y D (Mango)	13.00 a	13.40 a	0.72 e	0.68 f	262.1 a	274.3 a	72.26 f	67.02 d
C (water spray)	13.02 a	13.02 a-d	0.77 cde	0.77 cde	220.6 c	220.6 c	90.23 ab	90.23 a

AR: *Azospirillum* spray with 20 L / feddan; AD: *Azospirillum* spray with 40 L / feddan, SSC: soluble solids content.

Means with different letters within the same column differ significantly at P < 0.05.

On the other hand, the spray with *Azospirillum* isolates, generally, decreased color degree of fruits, and the differences than control were significant. Sahain *et al.* (2007) mentioned that the microbial spray with EM (Japanese inoculant) leads to improvement of most marketing characteristics of apple fruits except for firmness decrease. Pirlak and Köse (2009) claimed that spray of strawberry with PGPR (*Pseudomonas* BA-8) and *Bacillus* OSU-142 improved the quality characteristics of the fruits especially TSS.

Data of Table 5 illustrated the economical evaluation for the spray with different *Azospirillum* isolates, the treatments brought high increases in the net return per feddan. The highest net return was achieved by the application of the treatments of A¹R and A¹D that gave net return about 21870 and 21900 L E/fed. with an increase over net return of the control treatment by 6120 and 6190 L E/fed. respectively, followed by the spray with the treatments A²R and A²D which gave increase in net return over control by 0090 and 0480 L E/fed. respectively. The spray with bacterial biostimulants (*Azospirillum* isolates) resulted in considerable net return (L E/feddan), whereas, the application with the treatment A¹ increased the net return over control by 6120 L E/feddan. These results are in harmony with the results of Nour El-Din (2006) as the spray of peanut plants with liquid culture of *Azospirillum* lead to increase of the net return (L E/feddan).

Table 5: Response of "Anna" apple crop economics to spray with phyllosphere *Azospirillum* isolates

Treatments	Fixed costs (LE/ fed.)	Changed costs (LE / fed.)	Total costs (LE / fed.)	Total yield (Ton/ fed)	Crop value (LE /fed.)	Net return (LE /fed.)	Increase in return over control (LE)
A ¹ R (Lemone)	0...	6..	06..	7,63	26700	21100	0300
A ¹ D (Lemone)	0...	12..	62..	7,60	26700	20377	4622
A ² R (Guava)	0...	6..	06..	7,02	24070	18970	3210
A ² D (Guava)	0...	12..	62..	7,13	24900	18700	3000
A ¹ R (apple)	0...	6..	06..	7,80	27470	21870	6120
A ¹ D (apple)	0...	12..	62..	8,03	28100	21900	6190
A ² R (Orange)	0...	6..	06..	7,43	26000	20400	4600
A ² D (Orange)	0...	12..	62..	7,03	26300	20100	4400
A ¹ R(Mango)	0...	6..	06..	7,70	26900	21300	0090
A ¹ D (Mango)	0...	12..	62..	7,84	27440	21240	0480
C (water spray)	0...	0..	00..	0,93	20700	10700	-

AR: *Azospirillum* spray with 10 L / feddan; AD: *Azospirillum* spray with 20 L / feddan.

CONCLUSION

The foliar spray with bacterial biostimulants (*Azospirillum* isolates) increased apple growth and fruit yield, but the marketing quality of the fruits not significantly affected. The treatments were economically valuable. The used *Azospirillum* isolates were isolated from phyllosphere of different types of fruit trees (lemon, guava, apple, orange and mango), whereas the best efficient isolates was that isolated from apple leaves. The foliar application with PGPR biostimulants may become, in the near future, an effective tool for inducing growth and productivity of the plants.

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تقييم بعض عزلات الأروسبيرليم المنتخبة لإنتاج اندول حامض ألكليك وتأثيرها على تحسين نمو أشجار التفاح "صنف آنا" و محصول وجودة الثمار
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الرش الورقي بمزارع PGPR المخففة له أهمية قصوى لما تحتويه و تفرزه من مغذيات و منشطات و مضادات ميكروبية، ففقدرة هذه الميكروبات في منطقة الفيلاوسفير على تنشيط و تحسين نمو النبات بالإضافة إلى زيادة تحمل النبات للمرضات كان واضحا و مؤثرا. ولكن الأنواع الميكروبية بل و السلالات المختلفة تتباين في قدرتها على التأقلم و التعايش و إفراز المنشطات و المغذيات.

تم عزل ميكروبات الأروسبيرليم من أشجار مختلفة و هي التفاح و البرتقال و الليمون و الجوافة و المانجو و تم عمل مزارع نقية من هذه العزلات و استخدمت في رش أشجار تفاح بالجرعات ٢٠ و ٤٠ لتر للفدان في مقابل الرش بالماء كعقولة مقارنة. أظهرت النتائج ان الرش الورقي للأشجار بعزلات الأروسبيرليم قد زادت و بدرجة ملحوظة من نمو الأشجار و زادت إنتاجية الثمار ولم تؤثر معنويا على نوعية الثمار. ومع ذلك فقد اختلف تأثير و قدرة العزلات بدرجة معنوية، فقد كان التفوق في التأثير للعزلة A٣ تبعها A٧ و اللتان حققتا أعلى زيادة في طول المجموع الخضري و قطره و مساحة الورقة و وزنها الجاف و محتوى كلوروفيل أ و ب و المحتوى الكلي و كذلك إنتاجية الثمار في مقابل معاملة المقارنة، ولكن الخصائص النوعية للثمار (SSC و الحموضة و الصلابة و اللون) لم تظهر اختلافات معنوية ثابتة. وقد حققت معاملات الرش بعزلات الأروسبيرليم زيادة كبيرة في صافي العائد (بالجنيه المصري)، وكان في الصدارة معاملتنا الرش بالعزلة A٣ و D و A٣ و اللتان حققتا زيادة في صافي العائد عن معاملة المقارنة (الرش بالماء) بمقدار ٦١٢٠ و ٦١٩٥ جنيها على التوالي و تبعهما المعاملة A٧R (٥٥٩٥ جنيها/ فدان). وقد لوحظ أن الرش بتركيز ٤٠ لتر للفدان لم يعطي فروق معنوية عن الرش بتركيز ٢٠ لتر للفدان، وبالتالي فإننا نوصي بالرش بالعزلات الفعالة من الأروسبيرليم مثل العزلة A٣ و A٧ بالجرعة ٢٠ لترا للفدان، كذلك من المهم أن نركز الأبحاث على عزل و تقييم الميكروبات الفعالة لاستخدامها في التنمية الزراعية حيث أن لها عائد اقتصادي مجدي.

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